



## Phytochemical study of plants and plant cell cultures of three *Salvia* species

S. Schulz<sup>1</sup>, C. Haas<sup>1</sup>, S. Berkov<sup>2</sup>, A.I. Pavlov<sup>3,4</sup>, R. Ulber<sup>5</sup>, E. Neuhaus<sup>6</sup>, T. Bley<sup>1</sup>, J. Steingroewer<sup>1</sup>

<sup>1</sup>Institute of Food Technology and Bioprocess Engineering, TU Dresden, 01062 Dresden, Germany;

<sup>2</sup>AgroBioInstitute, 1164 Sophia, Bulgaria; <sup>3</sup>The Stephan Angeloff Institute of Microbiology, Bulgarian Academy of Sciences, 4000 Plovdiv, Bulgaria, <sup>4</sup>Department of Organic Chemistry and Microbiology, University of Food Technologies, 4002 Plovdiv, Bulgaria

<sup>5</sup>Institute of Bioprocess Engineering, 67663 University of Kaiserslautern, Germany; <sup>6</sup>Institute of Plant Physiology, University of Kaiserslautern, 67663 Kaiserslautern, Germany

### Introduction and Aims

The genus *Salvia* L. is widely distributed cultivated because of its numerous secondary metabolites with biological and pharmacological properties. Besides traditional production plant *in-vitro* cultures reveal a potential alternative source of selected secondary metabolites (see poster 1052, C. Haas).

For the screening of secondary metabolite production ethanolic callus extracts of three *Salvia* species (*S. triloba*, *S. officinalis* and *S. virgata*) have been analysed by GC-MS.

### GC-MS Analysis

Agilent GC 7890 MSD 5975C inert (EI 70eV)

HP-5ms column (30m x 250µm x 0.25µm)

He-Flow 1ml/min, injection 1µl, split: 1:50

Oven: 80°C (1min) with 10°C/min to 250°C then 2°C/min to 300°C (15min)

Internal standard cholesterole (c=104 µg/ml)

MSD simultaneous sim-scan-mode Scan 50-550m/z

Data analysis:

Comparison of measured mass spectra and RI-indices (C7-C40-Standard) after deconvolution with AMDIS 2.69 Software with Wiley 08, CSB DB Golm and GMD Golm Metabolome Database

Tab. 1 Detected metabolites and their biological and pharmaceutical properties

No.	Compound	Some biological and pharmacological properties
1	Trans-ferulic acid	antioxidative, anti-carcinogenic, antimicrobial
2	Trans-caffeic acid	antioxidative, anti-inflammatory, anti-carcinogenic
3	Stigmastan-3,5-diene	antioxidative, inhibits cholesterol absorption
4	β-Sitosterol	antioxidative, inhibits cholesterol absorption and proliferation of human leukemia cells
5	Rosmarinic acid	adstringent, antioxidative, antiplatelet, antibacterial, antiviral, antimutagen
6	Oleanolic acid (OA)	anti-inflammatory, antibacterial, hepatoprotective
7	Ursolic acid (UA)	anti-inflammatory, antibacterial, anticancerogen, antifungal, anti-tumor
8	3β-Acetoxy-oleanolic acid	anti-carcinogenic

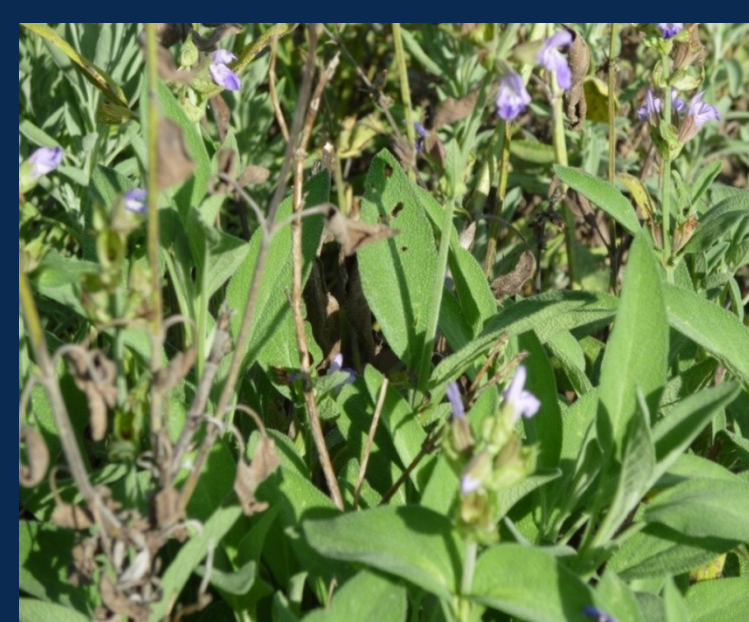


Fig. 1 *S. officinalis*



Fig. 2 callus of *S. officinalis*

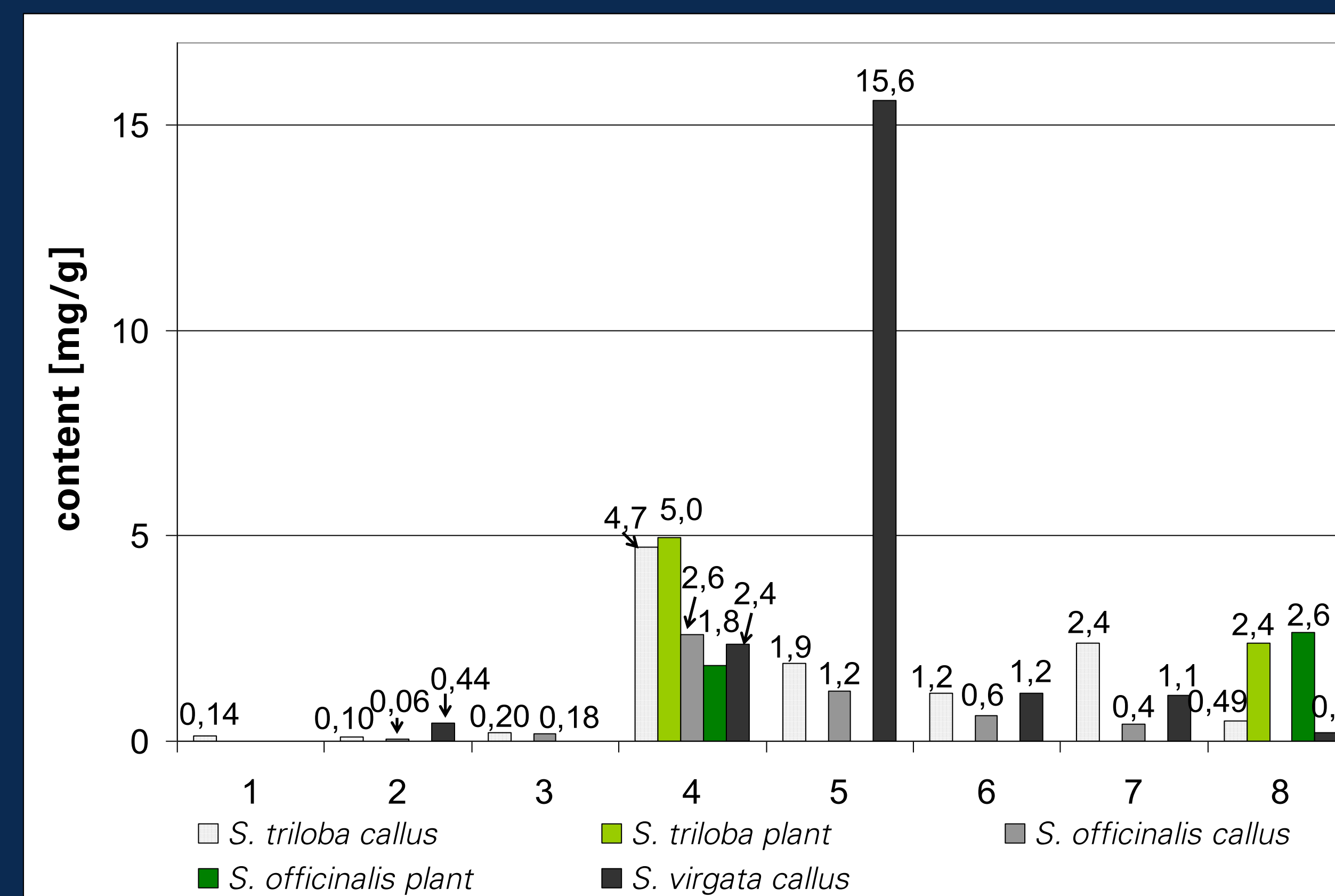


Fig. 3 Results of GC-MS screening of suspension cultures of *Salvia* sp., different cultivation phases and metabolites determined in plant material (numbering of the compounds see Tab. 1)

Determination of OA and UA in plant material by anionexchange chromatography as reference (recovery about 100%):

Tab. 2 Determined amount of OA and UA in plant material of *Salvia* sp.

Species	OA [mg/gDW]	UA [mg/gDW]
<b>S. triloba</b>	10.8	31.1
<b>S. officinalis</b>	7.5	22.4
<b>S. virgata</b>	5.5	2.8

### Conclusion and future prospects

The analysed callus extracts of the three *Salvia* species indicated the productivity of further pharmaceutical relevant metabolites: beside OA and UA also β-Sitosterol and Rosmarinic acid in high amounts, which deviates from the plants.

The productivity of OA and UA in *Salvia* suspension cultures (see poster 1052, C. Haas) is about 10% less compared with the plants and should be enhanced by cultivation optimisation and elicitation.

Further experiments for the isolation of OA and UA from the *in-vitro* cultures will be performed.